

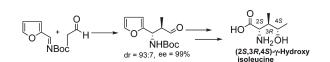
An Organocatalyzed Enantioselective Synthesis of (2*S*,3*R*,4*S*)-4-Hydroxyisoleucine and Its Stereoisomers

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A concise enantioselective total synthesis of (2S, 3R, 4S)-4-hydroxyisoleucine and its stereoisomers is described. A key feature of this protocol is a catalytic enantioselective mannich reaction that is either *anti*- or *syn*-selective as genesis of chirality.

The nonproteinogenic aminoacid (2S,3R,4S)-4-hydroxyisoleucine **2** has received renewed attention due to its broad range of pharmaceutical activities as insulinotropic, antidyslipidemic, and antihyperglycemic agent.¹ This unusual aminoacid was first isolated as free acid from fenugreek (*Trigonella foenum-graecum*) seeds² and the structure was validated by X-ray crystallography,³ establishing the absolute stereochemistry as 2S,3S,4R. Further, the SAR studies indicated that the absolute configuration of 2S,3R,4S stereogenic centers has considerable influence on their pharmaceutical activity (Figure 1).⁴

In fenugreek (2R,3R,4S)-4-hydroxyisoleucine *ent-3* was also found as a minor component. Later on, the (2S,3S,4R)-4-hydroxyisoleucine 3, which is a component of the natural product funebrine 4, was isolated from *Quararibea funebris* (Figure 1).⁵ Owing to the significant activity of these

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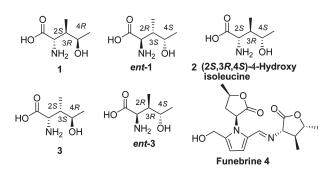


FIGURE 1. Stereoisomers of 4-hydroxyisoleucine.

molecules, impressive synthetic strategies have been reported. To date, most strategies rely on asymmetric induction resulting from either chiral auxiliaries⁶ resident chirality or enzymatic kinetic resolution of racemic mixtures.⁷

Herein, we report a highly practical and organocatalyzed enantioselective synthesis of (2S,3R,4S)-4-hydroxyisoleucine and its stereoisomers. In principle, the stereogenic centers 2 and 3 in (2S,3R,4S)-4-hydroxyisoleucine 2 could be accessed through a catalytic enantioselective Mannich reaction that are either anti-8 or syn-selective⁹ and by using N-Boc-furylimine 7 with 1-propanal 8. We chose furyl moiety as a masked carboxylic acid as well as to enhance its solubility in water.¹⁰ To obtain high selectivities (i.e., de) of the main product, we considered using water as the solvent in the Mannich reaction. Further, the stereogenic center 4 could be realized through a chelation-controlled nucleophilic addition of one carbon Grignard. Additionally, Mitsunobu inversion of stereogenic center 4 in 1, ent-1, 3, and ent-3 would generate a set of four diastereomers. Our retrosynthetic approach is shown in Scheme 1.

Accordingly, readily available furfural derived N-Bocimine 7 was treated with 8 in the presence of a catalytic

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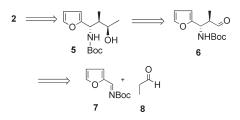
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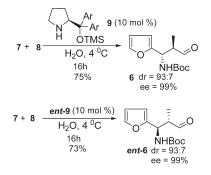
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⁽¹⁰⁾ The PMP-protected iminoesters have been used in organocatalyzed Mannich reactions. The protecting group PMP poses potential problems while deprotecting. The Boc-protected iminoesters were not suitable in our case. Hence, N-Boc-furylimine was employed in our study, which is easy to prepare and also hydrophilic in nature.

SCHEME 1. Retrosynthesis of (2*S*,3*R*,4*S*)-4-Hydroxy-isoleucine



SCHEME 2. TMS-Protected α, α -Diphenyl-2-pyrrolidinemethanol-Catalyzed Synthesis of β -Aminoaldehyde



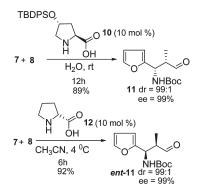
amount of TMS-protected α,α -diphenyl-2-pyrrolidinemethanol **9** (10 mol %) in water at 4 °C for 16 h. The desired β -aminoaldehyde **6** was isolated in 75% yield with 93:7 diastereoselectivity and 99% ee (Scheme 2).^{8a,11}

As anticipated, 10 mol % of *ent-9* under otherwise identical conditions resulted in *ent-6* in 73% yield with the same dr and ee. The diastereoselectivity was determined by integration of one set of ¹H NMR signals of the corresponding aldehyde (δ major 9.68 ppm as doublet and minor 9.75 ppm as singlet).¹¹ The ee value was analyzed by HPLC on the chiral staionary phase (Daicel Chiralpak OD-H column: 99/1 *n*-hexane/i-PrOH, flow rate 0.8 mL/min, major = 15.77 min).

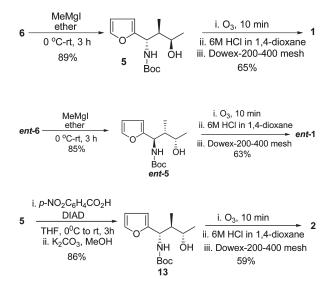
In a similar vein, L-proline-catalyzed *syn*-selective Mannich reaction^{9a} was evaluated employing **7** and **8** in water. To our surprise, no trace of the desired product **11** was isolated. When a similar reaction was conducted with 10 mol % of 4-*tert*-butyldiphenylsilyloxy L-proline, the required Mannich adduct **11** was isolated in 89% yield with a dr value of 99:1¹¹ and 99% ee.

However, the *syn*-variant *ent*-11 was generated with a catalytic amount of (*R*)-proline 12 (10 mol %) in acetonitrile at 4 °C. The resulting product *ent*-11 was isolated in good yield (92%) with high diastereo-enantioselectivity (dr 99:1, ee >99%) (Scheme 3).¹¹ With *anti*-*syn*-varients of β -aminoaldehydes in hand, we examined the addition of one carbon Grignard for the synthesis of β -aminoalcohols. The exposure of **6** to 1.5 equiv of methylmagnesium iodide at 0 °C to rt for 3 h furnished **5** in 89% yield as the only isolable diastereomer. In the same way, *ent*-**6** subjected to identical conditions (vide infra) resulted in *ent*-**5** in 85% yield with the same optical purity. The diastereoselectivity could be rationalized on the basis of a chelation-controlled mechanism wherein the nucleophile is approaching from the *re-face* of

SCHEME 3. Synthesis of syn-Varient β -Aminoaldehydes



SCHEME 4. Synthesis of (2*S*,3*R*,4*R*)-4-Hydroxyisoleucine and Its Stereoisomers



the pro-carbonyl group leading to the observed product **5**, while the adjacent methyl-substituted stereogenic center does not play any significant role.

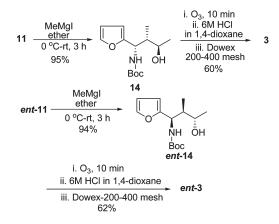
Oxidative cleavage of the furyl moiety of **5** (O₃, 10 min) followed by Boc deprotection under acidic conditions (6 M HCl in dioxane) and subsequent purification (Dowex 50WX8-400) led to (2S,3R,4R)-4-hydroxyisoleucine **1** in 65% yield. Inversion of stereogenic center **4** in **5** under Mitsunobu conditions (*p*-NO₂C₆H₄CO₂H, DIAD, THF, rt, 3 h/K₂CO₃, MeOH) afforded **13** in 86% yield.

Compound 13 was subjected to oxidation, followed by deprotection of the Boc group and purification resulting in the title compound 2 with 59% isolated yield. The chemical data of 2 are identical with those reported for the naturally occurring molecule ($[\alpha]^{24}_{D}$ +31.2 (*c* 0.9, H₂O), lit.² $[\alpha]^{24}_{D}$ +31 (*c* 1, H₂O)) (Scheme 4). As a result, the absolute stereo-chemistry of 2 was assigned as 2*S*,3*R*,4*S*, which inturn conformed with the absolute configuration of 1 as 2*S*,3*R*,4*R*.

The above-mentioned addition of Grignard/oxidation/ Boc-deprotection was repeated with compound 14 and *ent*-14 to give 3 and *ent*-3 in 60% and 62% yields, respectively. The relative stereochemistry of 3 was assigned as 2S,3S,4Rbased on single X-ray crystallography (CCDC reference no. 764863;¹² the crystal was obtained from 5% EtOAc in hexane (see the Supporting Information)). The optical rotation

⁽¹¹⁾ The dr values of **6**, *ent*-**6**, **11**, and *ent*-**11** have been determined after chromatography of the corresponding reaction mixture (see the Supporting Information).

SCHEME 5. Synthesis of (2S, 3R, 4R)-4-Hydroxyisoleucine and Its Stereoisomer



of *ent*-14 was found to be approximately equal in magnitude to that of 14 but opposite in sign, indicating an enantiomeric relationship, hence stereochemistry of *ent*-3 was assigned as 2R, 3R, 4S (Scheme 5).

In conclusion, we have accomplished a concise enantioselective total synthesis of (2S,3R,4S)-4-hydroxyisoleucine **2** and its stereoisomers. Strategic transformation includes a catalytic enantioselective Mannich reaction that is either *syn*- or *anti*-selective as genesis of chirality, methyl Grignard addition, and Mitsunobu inversion to generate eight stereoisomers with perfect stereocontrol. To our knowledge, no catalytic diastereo-enantioselective variant reaction has been explored before for the synthesis of (2S,3R,4S)-4hydroxyisoleucine. Moreover, flexibility was built into the synthesis to generate a library of analogues. This protocol is also amenable to large-scale synthesis of nonproteinogenic aminoacid.

Experimental Section

tert-Butyl (1S,2R)-1-(Furan-2-yl)-2-methyl-3-oxopropylcarbamate (6). 1-Propanal 8 (594 mg, 10.25 mmol) and N-Bocprotected imine 7 (1.0 g, 5.12 mmol) were added to a roundbottomed flask charged with catalyst 9 (167 mg, 10 mol %) and 2 mL of H₂O at 4 °C. The reaction mixture was stirred for 16 h at this temperature, then the reaction was quenched by addition of EtOAc (15 mL) and the mixture was extracted with EtOAc (3 \times 10 mL). The organic layer was separated and dried over Na₂SO₄, concentrated, and evaporated to give crude product. The crude residue was subjected to column chromatography eluting with hexane/EtOAc (95/5) furnishing 6 as a colorless liquid (973 mg, 75%) with dr 93:7 and 99% ee [dr 93:7, determined by integration of one set of ${}^{1}H$ NMR signals $(\delta_{\text{major}} 9.65 \text{ ppm, d}; \delta_{\text{minor}} 9.73 \text{ ppm, s})]$. HPLC analysis on a DaicelChiralpak OD-H column: 99/1 n-hexane/i-PrOH, flow rate 0.8 mL/min, $\lambda = 215$ nm; $\tau_{major} = 15.77$ min; $[\alpha]^{24}_{D} + 19.8$ (c 0.9, CHCl₃, 99% ee). IR (KBr) 3365, 2979, 2932, 1728, 1681, 1527, 1451, 1372, 1276, 1169, 1051, 919, 753, 612 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 9.73 (1H, s), 7.34–7.31 (1H, m), 6.30-6.28 (1H, m), 6.20-6.18 (1H, m), 5.21-5.01 (2H, m), 2.97-2.84 (1H, m), 1.44 (9H, s), 1.09 (3H, t, J = 7.5 Hz); ¹³C NMR (150 MHz, CDCl₃) δ 202.2, 141.6, 109.8, 106.8, 49.4, 27.7, 9.1; MS (ESIMS) m/z 254 (M + H)⁺, 276 (M + Na)⁺; HRMS (ESI) m/z 276.1215 (calcd for C₁₃H₁₉NO₄Na 276.1211).

tert-Butyl (1S,2R,3R)-1-(Furan-2-yl)-3-hydroxy-2-methylbutylcarbamate (5). An ether solution of aldehyde 6 (450 mg, 1.78 mmol, 20 mL) was added dropwise to a cooled (0 °C) solution of methylmagnesium iodide prepared from magnesium (64 mg, 2.67 mmol) and methyl iodide (336 mg, 2.67 mmol) in dry ether (15 mL). After addition, the resulting solution was stirred at room temperature for 3 h. The solution was then slowly poured into crushed ice, and the precipitated magnesium hydroxide was quenched by the addition of saturated ammonium chloride (30 mL). The organic layer was separated, and the aqueous phase was saturated with sodium chloride (20 mL) and extracted with chloroform $(3 \times 15 \text{ mL})$. The combined organic layers were dried over Na₂SO₄ and filtered. The solvent was removed under reduced pressure. The crude residue was subjected to column chromatography eluting with hexane/EtOAc (90/10) to furnish 5 as a colorless liquid (425 mg, 89%). $[\alpha]^{24}{}_{\rm D}$ +25.8 (*c* 0.9, CHCl₃). IR (KBr) 3424, 2278, 2972, 2931, 1675, 1542, 1504, 1365, 1267, 1168, 1092, 1009, 935, 748, 695 cm⁻¹; ¹H NMR (300 MHz, $CDCl_3$) δ 7.34–7.33 (1H, m), 6.31–6.29 (1H, m), 6.16 (1H, d, J = 3.2 Hz), 4.91-4.85 (2H, m), 3.82-3.72 (1H, m), 1.97-1.88 (1H, m), 1.46(9H, s), 1.21(3H, d, J = 6.2 Hz), 0.95(3H, t, J = 6.8 Hz); ¹³C NMR (150 MHz, CDCl₃) δ 141.2, 109.9, 106.8, 69.2, 50.3, 45.5, 28.1, 20.2, 11.8; MS (ESIMS) m/z 270 (M + H)⁺, 292 (M + Na)⁺; HRMS (ESI) *m*/*z* 292.1533 (calcd for C₁₄H₂₃NO₄Na 292.1524).

tert-Butyl (1S,2R,3S)-1-(Furan-2-yl)-3-hydroxy-2-methylbutylcarbamate (13). Triphenylphosphine (780 g, 2.98 mmol), p-nitrobenzoic acid (279 g, 1.48 mmol), and compound 5 (400 mg, 1.48 mmol) were dissolved in THF (10 mL). To this mixture was added a solution of diisopropylazodicarboxylate (601 mg, 2.97 mmol) in THF (5 mL) at 0 °C via a syring. The reaction contents were stirred at room temperature. After 3 h, the reaction mixture was concentrated under vacuum. The crude residue was dissolved in methanol (20 mL) and cooled to 0 °C. To this was added potassium carbonate (410 mg, 2.97 mmol) portionwise. The mixture was warmed to room temperature over a period of 1 h. Then the solvent was removed under vacuum and the residue was dissolved in CH₂Cl₂ (20 mL). The organic layer was washed with brine (20 mL) and then separated. The aqueous layer was extracted with CH_2Cl_2 (2 × 10 mL), and the combined organic layers were dried over Na₂SO₄ and concentrated. The crude residue was subjected to silica gel column chromatography (100-200 mesh), using hexane and ethyl acetate (90:10) as solvents, yielding the pure product 13 as a colorless liquid (344 mg, 86%). $[\alpha]^{24}_{D}$ +28.8 (c 0.9, CHCl₃). IR (KBr) 3414, 2281, 2977, 2932, 1678, 1547, 1517, 1366, 1270, 1173, 1092, 1012, 965, 745, 689 cm⁻¹; 1 H NMR (300 MHz, CDCl₃) δ 7.32–7.31 (m, 1H), 6.29–6.31 (m, 1H), 6.19 (1H, m), 5.14–5.12 (1H, m), 4.96–4.84 (1H, m), 3.64-3.62 (1H, m), 1.20-1.98 (1H, m), 1.46 (9H, s), 1.98 (3H, d, J = 6.3 Hz), 0.80 (3H, t, J = 6.9 Hz); ¹³C NMR (150 MHz, CDCl₃) & 142.0, 110.7, 106.3, 69.0, 51.9, 43.5, 28.5, 21.1, 8.8; MS $(\text{ESIMS}) m/z 270 (M + H)^+, 292 (M + Na)^+; \text{HRMS} (\text{ESI}) m/z$ 292.1529 (calcd for C₁₄H₂₃NO₄Na 292.1524).

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⁽¹²⁾ The crystallographic coordinates have been deposited with the Cambridge Crystallographic Data Centre; deposition no. 764863. These data can be obtained free of charge from the Cambridge Crystallographic Data Centre, 12 Union Rd., Cambridge CB2 1EZ, UK or via www.ccdc.cam. ac.uk/conts/retrieving.html.

Supporting Information Available: Experimental procedures and characterization data for all new compounds along with copies of ¹H and ¹³C NMR spectra, and crystallographic data. This material is available free of charge via the Internet at http://pubs.acs.org.